A New Benzyol Compound Isolated from the Endophytic Fungi of Kandis Gajah (Garcinia griffithii) and Asam Kandis (Garcinia cowa)

Elfita*, Munawar2, Muharni1, Gusti Pratiwi1, and Rahmadania1

1. Department of Chemistry, Faculty of Mathematics and Natural Sciences, Universitas Sriwijaya, Indralaya, Kabupaten Ogan Ilir 30662, Indonesia
2. Department of Biology, Faculty of Mathematics and Natural Sciences, Universitas Sriwijaya, Indralaya, Kabupaten Ogan Ilir 30662, Indonesia

*E-mail: elfita_kimia@unsri.ac.id
Received March 11, 2015 | Accepted February 22, 2016

Abstract

Garcinia griffithii and Garcinia cowa belong to the genus Garcinia. The genus Garcinia has been known to be a rich source of secondary metabolites, such as xanthones, benzophenones, flavonoids, steroids, terpenoids, and other phenolic derivatives. Previous investigations of endophytic fungi from G. griffithii revealed the presence of three compounds not found in the host. In order to continue the phytochemical work on endophytic fungi of G. griffithii, the constituent of the endophytic fungi of G. griffithii was re-examined. In this study, a benzyol compound similar to that found in the endophytic fungus of G. cowa was observed. The same benzyol compound was also isolated from the endophytic fungus Acremonium sp of G. griffithii and Aspergillus sp of G. cowa with cultivation of eight weeks in static conditions at room temperature. The culture medium was partitioned using ethyl acetate and evaporated to obtain the concentrated extract. Isolation of compounds was performed using the chromatography method. The chemical structure was proposed on the basis of spectroscopic data, including ultraviolet (UV), infrared (IR), mass spectrometry (MS), proton nuclear magnetic resonance (1H-NMR), carbon nuclear magnetic resonance (13C-NMR), heteronuclear single-quantum correlation spectroscopy (HSQC), heteronuclear multiple-bond correlation spectroscopy (HMBC), and correlation spectroscopy (COSY).

Abstrak

Suatu Senyawa Benzoil Baru Hasil Isolasi dari Jamur Endofit Tumbuhan Kandis Gajah (Garcinia griffithii) dan Asam Kandis (Garcinia cowa). Tumbuhan Garcinia griffithii dan Garcinia cowa termasuk ke dalam genus Garcinia. Genus Garcinia dikenal kaya dengan kandungan metabolit sekunder, seperti santon, benzofenon, flavonoid, steroid, terpenoid, dan turunan fenolik lainnya. Penelitian sebelumnya telah menemukan tiga senyawa yang tidak ditemukan dalam tumbuhan inangnya. Sebagai lanjutan dari penelitian atau kandungan kimia dari jamur endofit ini, penelitian ini diperoleh benzoil yang sama dengan yang ditemukan dalam jamur endofit tumbuhan G. Cowa. Senyawa benzoil yang sama juga dapat diisolasi dari jamur Acremonium sp tumbuhan G. griffithii dan dari jamur Aspergillus sp tumbuhan G. cowa dengan masa kultivasi delapan minggu dalam kondisi statis pada suhu kamar. Mempelajari pertumbuhan diperoleh esensi dan dievaporasi untuk mendapatkan ekstrak pekatnya. Isolasi senyawa dilakukan dengan metode kromatografi. Struktur kimia diusulkan berdasarkan data spektroskopi yang meliputi ultraviolet (UV), infrared (IR), mass spectrometry (MS), proton nuclear magnetic resonance (1H-NMR), carbon nuclear magnetic resonance (13C-NMR), heteronuclear single-quantum correlation spectroscopy (HSQC), heteronuclear multiple-bond correlation spectroscopy (HMBC), dan correlation spectroscopy (COSY).

Keywords: benzyol, endophytic fungi, Garcinia griffithii, Garcinia cowa

Introduction

Endophytic fungi are a new source of bioactive compounds being explored recently. They live in plant tissue in a certain period of time, can form colonies in plant tissues without harming the host, and usually provide symbiotic mutualism [1-3]. One of the interesting facts about endophytic fungi is their ability to produce bioactive compounds that can be similar or different with those produced by the host [4-5].
Plants of the genus *Garcinia* are known as a rich source of diversity for secondary metabolites including xanthones, benzophenone, flavonoids, steroids, terpenoids, and other phenolic derivatives [6-8]. The extract of natural products have been used as drugs and important sources of traditional medicines, such as cytotoxic, antibacterial, antiviral, antifungal, antioxidant, anti-inflammatory, antidepressant, and anti-HIV [9-11].

Phongpaichit *et al.* [2007] and Ruma *et al.* [2013] reported endophytic fungal isolated from the *Garcinia* genus, including *Garcinia atroviridis*, *Garcinia dulcis*, *Garcinia mangostana*, *Garcinia nigrolinnea*, *Garcinia Scortechinii*, *Garcinia gummigutta*, *Garcinia indica*, *Garcinia morella*, and *Garcinia xanthochymus*. The endophytic fungi are shown to have antimycobacterial, antimalarial, antiviral, antioxidant, anti-inflammatory, and cytotoxicity activities [12-13]. Several strategies used to isolate endophytic fungi have been reported recently to search for potential bioactive compounds, and one of them is based on ethnobotanical history specifically used by indigenous peoples [14]. *Garcinia griffithii* and *Garcinia cowa* are traditionally used by the local communities of Sarasar Bonta, Lemhah Arau, and West Sumatra to treat various diseases, including gout, diarrhea, and malaria.

In previous work, we reported three compounds from endophytic fungi of *G. griffithii*, namely, 4,6-dihydroxy-3,8a-dimethyl-1-oxo-5-(3”-oxobutan-2”-yl)-1,4, 4a, 5, 6, 8a-hexahydroaphthalen-2-yl-1”, 2”-di-methyl- 5”-(2”’-methyl-prop-1”’-eny)-cyclopentan-carboxylate from *Aspergillus fumigatus* [15]; 3,5-dihydroxy-2,5-dimethyltrideca-2,9,11-triene-4,8-dione from *Acremonium* sp [16]; and 10, 12-trihydroxy-9-methoxy-7a-methyl-7, 7a, 12a, 13- tetrahydrobenzo-cyclohepta-oxocin-6-one from *Aspergillus niger* [17].

In the current study, we reported another secondary metabolite called benzoyl compound, which is extracted from *Acremonium* sp of *G. griffithii* and found in the endophytic fungus *Aspergillus* sp of *G. cowa*.

**Materials and Methods**

**Source of endophytic fungi.** *Acremonium* sp of *G. griffithii* obtained from stock fungus was stored in the Microbiology Laboratory, Department of Biology, Faculty of Mathematics and Natural Sciences of Sriwijaya University. The fruit of *G. cowa* was collected in May 2012 from Lemhah Arau, West Sumatra.

**Isolation and identification of endophytic fungi.** The isolation of *Acremonium* sp from twigs of *G. griffithii* was reported previously by Elfita *et al.* [2012] [16] and Debbab *et al.* [18]. The isolation method of *Aspergillus* sp from the fruit of *G. cowa* grown in Sarasar Bonta, Lemhah Arau, Kabupaten Lima Pulu Kota, and West Sumatra was conducted according to the procedure by Elfita *et al.* 2011 [15]. The fungal strain was identified on the basis of the morphological method by the School of Hayati Science and Engineering, Bandung Institute of Technology, Indonesia. The voucher of specimen was stored in the Microbiology Laboratory, Department of Biology, Faculty of Mathematics and Natural Sciences of Sriwijaya University.

**Cultivation of endophytic fungi.** Potato dextrose broth (PDB) medium of 300 mL was used for the cultivation of endophytic fungi and was placed into 30 flasks (1 L each). Fungal suspension containing 10⁵ spores/mL was inoculated under sterile conditions to each 300 mL PDB medium (ratio 1:10). The cultures were incubated for eight weeks in static conditions at room temperature [19-20].

**Extraction, exploration, and structure elucidation.** Mycelia were removed from the endophytic fungus culture after eight weeks of incubation and the medium was filtered. The medium was extracted three times using ethyl acetate (1:1) followed by evaporation under vacuum to obtain the concentrated extract. The concentrated extract was separated by column chromatography over silica gel 60 (70–230 mesh) at the stationary phase (1:30) and eluent, which was previously determined by thin-layer chromatography silica gel 60 F254. The chosen eluent with increased polarity was n-hexane:EtOAc at a ratio of 10:0 to 10:10 (v/v). An eluate was collected and then combined using thin-layer chromatography into column fractions. Each fraction was evaporated and purified using the chromatography technique to obtain the purified compound.

The ethyl acetate extract from *Acremonium* sp of *G. griffithii* (3.16 g) was subjected to column chromatography over silica gel (60 x 1.5 cm) and was eluted with n-hexane-ethyl acetate gradient (10:0–0:10) and collected in 60 vials each containing 10 mL. The thin layer chromatography (TLC) analysis showed the presence of five column fractions (F1–F5): F1 (724 mg), F2 (433 mg), F3 (406 mg), F4 (821 mg), and F5 (891 mg). Fraction F3 showed a major compound. It was further separated by column chromatography over silica gel (30 x 0.7 cm) and eluted with n-hexane-ethyl acetate (5:5–0:10) to give 37 vials. The fractions, which gave the same *Rf* on TLC, were combined and yielded four column fractions (F3.1–F3.4); F3.1 (81 mg), F3.2 (52 mg), F3.3 (143 mg), and F3.4 (105 mg). Fraction F3.3 was purified by rechromatography to yield compound 1 as a white crystal (87 mg). Its melting point is 109 °C–111 °C. Further separation and purification of fraction F4 by column chromatography yielded compound 2 as previously reported by Elfita *et al.* 2012 [16].

The ethyl acetate extract from *Aspergillus* sp of *G. cowa* (3.02 g) was subjected to column chromatography on silica gel (60 x 1.5 cm), eluted with n-hexane-ethyl acetate gradient (10:0–0:10), and collected in 50 vials each containing 10 mL. The TLC analysis showed the

Makara J. Sci.
The fungal strain from the fruit of *G. cowa* was identified as *Aspergillus* sp by the School of Hayati Science and Engineering, Bandung Institute of Technology, Indonesia. *Aspergillus* sp was cultivated on 9 L of PDB medium for eight weeks at room temperature. The culture broth was extracted by solvent partition with EtOAc (1:1), followed by evaporation. The extract showed one major spot on TLC. Figure 1 briefly illustrates the procedure for isolating the pure compound from the endophytic fungi. *Acremonium* sp from the twigs of *G. griffithii* and *Aspergillus* sp from the fruit of *G. cowa*.

**Figure 1.** Brief Procedure for Isolating the Pure Compound from the Endophytic Fungi *Acremonium* sp from Twigs *G. griffithii* and *Aspergillus* sp from the Fruit of *G. cowa*
The presence of five aromatic protons showed the signal at δH 7.48 ppm (2H, d, J=7.8), 8.32 ppm (2H, d, J=7.8), and 7.64 ppm (1H, t, J=7.5). A signal was also found for methoxy proton at δH 3.45 ppm (3H, s) and two vinylic protons at δH 5.56 ppm (1H, m) and 5.23 ppm (1H, m). The 1H-NMR revealed the presence of signals to three methylene protons sp2 as oxygenated carbon at δH 4.69 ppm (1H, s), 4.74 ppm (1H, dd, J=4.5; 3.9), and 4.59 ppm (1H, d, J=3.9).

The 13C-NMR spectrum showed 22 signals, including 8 carbon sp2 and 14 carbon sp3. The 13C-NMR and DEPT 135 spectrum, which were supported by HSQC spectrum, showed eight carbon sp3 signals consisting of two methyl, one methylene, three methine, one methoxy, and one quaternary carbons.

Three signals of methine at δC 70.8, 71.3, and 73.1 ppm were assigned to carbon atoms bearing hydroxyl groups and one quaternary carbon bearing a methoxyl group. Furthermore, 14 carbon sp3 signals included 6 aromatic carbons at δC 128.9 and 135.0, and 2 carbons with the same chemical shift at 131.0 ppm and 132.6 ppm. Two of these signals were assigned to the ketone carbons at δC 195.6 ppm and 196.5 ppm. Six other sp2 carbon signals were present at δC 93.1, 113.3 126.7, 136.7, 166.7, and 186.2 ppm.

In the HMBC spectrum (Table 1.), correlations from H-3'' (δH 8.32) to C-1'' (δC 195.6), C-4'' (δC 128.9), and C-5'' (δC 135.0) suggested that C-1'' was attached to the aromatic ring as a benzyl group. Correlations from the hydroxy proton at C-3' (δH 8.46) to C-2' (δC 93.1) and C-4' (δC 73.1) and from 1'-OCH3 (δH 3.45) to C-1' (δC 90.6) demonstrated the presence of a cyclobutene ring bearing methoxyl and hydroxy groups. The methoxyl resonance connecting to C-1' in HMBC indicated C-1' with a high chemical shift (δC 90.6). Moreover, oxygenated carbon was attached to C-1 (carbonyl carbon). Correlations from the hydroxy proton at C-3' (δH 8.46) to C-2' with a low chemical shift as sp2 C=C (δC 93.1) showed an anisotropy effect of carbonyl groups (C-1').

HMBC correlations from 2-CH3 (δH 1.66) to C-1 (δC 196.5), C-2 (δC 113.3), and C-3 (δC 186.2) indicated that C-1 and C-3 were located at the carbonyl group and hydroxyl group, respectively. Correlations from H-4 (δH 4.59) to C-5 (δC 71.3) and H-5 (δH 4.74) to C-4 (δC 70.8) revealed that C-4 and C-5 were attached to the hydroxyl groups. Figure 2 illustrates the HMBC correlation and δ-assignment of compound I.

**Table 1. NMR Data of Compound 1 recorded at 1H-500MHz, 13C-125 MHz in CDCl3**

<table>
<thead>
<tr>
<th>Position of C</th>
<th>δC ppm</th>
<th>δH ppm</th>
<th>ΣH, multiplicity, J (Hz)</th>
<th>HMBC</th>
<th>COSY</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>196.5</td>
<td>196.5</td>
<td>H-5</td>
<td></td>
<td></td>
</tr>
<tr>
<td>2</td>
<td>113.3</td>
<td>113.4</td>
<td>H-5</td>
<td></td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>186.2</td>
<td>186.1</td>
<td>H-5</td>
<td></td>
<td></td>
</tr>
<tr>
<td>4</td>
<td>70.8</td>
<td>70.8</td>
<td>4.59 (1H;d; 3.9)</td>
<td>C-8</td>
<td>C-8</td>
</tr>
<tr>
<td>5</td>
<td>71.3</td>
<td>71.2</td>
<td>4.74 (1H;m)</td>
<td>C-9</td>
<td>C-9</td>
</tr>
<tr>
<td>6</td>
<td>126.7</td>
<td>126.7</td>
<td>5.12 (1H;m)</td>
<td>C-10</td>
<td>C-10</td>
</tr>
<tr>
<td>7</td>
<td>136.7</td>
<td>136.8</td>
<td>5.56 (1H;m)</td>
<td>C-11</td>
<td>C-11</td>
</tr>
<tr>
<td>8</td>
<td>21.6</td>
<td>21.6</td>
<td>2.11 (2H;m)</td>
<td>C-12</td>
<td>C-12</td>
</tr>
<tr>
<td>9</td>
<td>14.3</td>
<td>14.3</td>
<td>0.97 (3H;t;7.2)</td>
<td>C-13</td>
<td>C-13</td>
</tr>
<tr>
<td>1'</td>
<td>90.6</td>
<td>90.6</td>
<td>H-6</td>
<td></td>
<td></td>
</tr>
<tr>
<td>2'</td>
<td>93.1</td>
<td>93.0</td>
<td>H-6</td>
<td></td>
<td></td>
</tr>
<tr>
<td>3'</td>
<td>166.7</td>
<td>166.9</td>
<td>H-6</td>
<td></td>
<td></td>
</tr>
<tr>
<td>4'</td>
<td>73.1</td>
<td>73.2</td>
<td>4.69 (1H,s)</td>
<td>C-14</td>
<td>C-14</td>
</tr>
<tr>
<td>1'*</td>
<td>195.6</td>
<td>195.5</td>
<td>H-6</td>
<td></td>
<td></td>
</tr>
<tr>
<td>2'*</td>
<td>132.6</td>
<td>132.5</td>
<td>H-6</td>
<td></td>
<td></td>
</tr>
<tr>
<td>3'*</td>
<td>131.0</td>
<td>131.0</td>
<td>8.32 (2H;7.8)</td>
<td>C-15</td>
<td>C-15</td>
</tr>
<tr>
<td>4'*</td>
<td>128.9</td>
<td>128.9</td>
<td>7.48 (2H;m)</td>
<td>C-16</td>
<td>C-16</td>
</tr>
<tr>
<td>5'*</td>
<td>135.0</td>
<td>135.0</td>
<td>7.64 (1H;t;7.5)</td>
<td>C-17</td>
<td>C-17</td>
</tr>
<tr>
<td>6'*</td>
<td>128.9</td>
<td>128.9</td>
<td>7.48 (2H;m)</td>
<td>C-18</td>
<td>C-18</td>
</tr>
<tr>
<td>7'*</td>
<td>131.0</td>
<td>131.0</td>
<td>8.32 (2H;7.8)</td>
<td>C-19</td>
<td>C-19</td>
</tr>
<tr>
<td>2-CH3</td>
<td>6.3</td>
<td>6.3</td>
<td>1.66 (3H)s</td>
<td>C-20</td>
<td>C-20</td>
</tr>
<tr>
<td>1'-OCH3</td>
<td>51.9</td>
<td>51.9</td>
<td>3.45 (3H)s</td>
<td>C-21</td>
<td>C-21</td>
</tr>
<tr>
<td>3'-OH</td>
<td>8.46</td>
<td>8.46</td>
<td>8.46 (1Hbr)</td>
<td>C-22</td>
<td>C-22</td>
</tr>
</tbody>
</table>

1: Compound I from Acremonium sp from the twig of G. griffithii
1*: Compound I from Aspergillus sp from the fruit of G. cowa

---

Maeka J. Sci.  
December 2016 | Vol. 20 | No. 4
In the $^1$H-$^1$H COSY spectrum, the correlations from H-4 to H-5 indicated the presence of vicinal hydroxyl groups. The next correlations from H-6 to H-5, H-7 and from H-8 to H-9 revealed the presence of long-chain aliphatic subunits. The correlation of $^1$H-$^1$H aromatic indicated a mono-substituted aromatic ring. The COSY correlation of compound 1 is illustrated in Figure 2.

Carbon C-1’ of compound 1 had a similar position and chemical environment to C-1’ of talaroflavone [21]. Carbon C-1’of the two compounds is C sp3 (quarterner), which binds two C sp2, one oxygenated C sp3, and one oxygen atom. Each carbon appeared at $\delta$C 90.6 and 91.8 ppm. Furthermore, carbon C-2’ of compound 1 had a position and chemical environment similar to those of C-3 of methyltriaceitic lactone [21], each of which appeared on $\delta$C 93.1 and 95.0 ppm. The chemical shift of C-1’ ($\delta$C 90.6) was shown to approach the calculated data using Chem Draw Ultra 10 ($\delta$C 94.8).

Combining the results obtained by the spectrometric methods (UV, IR, MS, $^1$H-NMR, $^1$C-NMR, HSQC, HMBC, and COSY) and comparing the results of the literature data [21] with the calculated data of Chem Draw Ultra 10, we determined C$_2$H$_3$O$_6$ as the molecular formula for compound 1 with a molecular weight of 418. The structure of compound 1 is 1-(2’-benzoyl-3,4-dihydroxy-1’-methoxycyclobut-2’-enyl)-3, 4, 5-trihydroxy- 2-methylnona-2,6-dien-1-one. The confirmation structure of 1 will be followed by an X-ray single crystal, and this work is still in progress.

Elucidation of compound 1* from Aspergillus sp of G. cowa. The $^1$H-NMR spectra recorded in CDCl$_3$ ($^1$H-500 MHz) indicated that the spectroscopy data of 1 and 1* are identical (see Table 1).

Based on the Dictionary Natural Products database (December 18, 2015), the benzoyl derivative could be a new compound. However, the identical benzoyl compounds (1 and 1*) were produced by two different endophyte fungi could further raise the question of whether it is related to the host of the same genus or assumed as a secondary metabolite typically produced by the fungi. Based on these phytochemical studies, further studies on the metabolite profiling of these endophytic fungi and plants should be performed for confirmation.

Conclusions

The endophytic fungi *Acremonium* sp from the twigs of *G. griffithii* produced the newly proposed benzoyl compound, which is identical to that found from the endophytic fungus *Aspergillus* sp from the fruit of *G. cowa*.

Acknowledgements

The authors are grateful to the Directorate General of Higher Education for funding this research through the Strategis Nasional Grant in 2011 entitled “Produksi Sediaan Obat Tradisional Terstandar untuk Asam Urat dari Mikroba Endofitik Tumbuhan Kandis Gajah (*Garcinia griffithii* T. Anders)” with contract No. 397/SP2H/PL/Dit.Litabmas/IV/2011.

References


